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☐	L14	starburst dendrimer and (PLGA! or polylactic-co-glycolic or polylcatide-co-glycolide)	7
☐	L13	starburst dewndrimer and (PLGA! or polylactic-co-glycolic or polylcatide-co-glycolide)	0
☐	L12	dendrimer and dna and (PLGA! or polylactic-co-glycolic or polylcatide-co-glycolide)	34
☐	L11	l7 and l2	6
☐	L10	5338532.pn.	2
☐	L9	l6 and l2	32
☐	L8	L7 and l2	6
☐	L7	membrane same (PLGA! or polylactic-co-glycolic or polylcatide-co-glycolide)	68
☐	L6	membrane same (polyester or lactide or glycolide or lactic or glycolic or bioerodible or biocompatible)	12105
☐	L5	L4 and l3 and l2	26
☐	L4	biological agent	3883
☐	L3	tissue	378604
☐	L2	dendrimer	2810
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1. **Michael addition reactions in macromolecular design for emerging technologies** • REVIEW ARTICLE
Progress in Polymer Science, Volume 31, Issue 5, May 2006, Pages 487-531
Brian D. Mather, Kalpana Viswanathan, Kevin M. Miller and Timothy E. Long
SummaryPlus | Full Text + Links | PDF (1307 K)

2. **BIOMATERIALS AND GENE THERAPY** • REVIEW ARTICLE
Advances in Chemical Engineering, Volume 29, 2004, Pages 131-168
F. Kurtis Kasper and Antonios G Mikos
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3. **The use of synthetic polymers for delivery of therapeutic antisense oligodeoxynucleotides** • REVIEW ARTICLE
Biomaterials, Volume 23, Issue 2, January 2002, Pages 321-342
Traian V. Chirila, Piroska E. Rakoczy, Kerryn L. Garrett, Xia Lou and Ian J. Constable
SummaryPlus | Full Text + Links | PDF (280 K)

4. **Plasmid DNA encapsulation and release from solvent diffusion nanospheres** • ARTICLE
Journal of Controlled Release, Volume 70, Issues 1-2, 29 January 2001, Pages 231-242
S. Hirosue, B. G. Müller, R. C. Mulligan and R. Langer
SummaryPlus | Full Text + Links | PDF (806 K)

5. **The delivery of antisense therapeutics** • ARTICLE
Advanced Drug Delivery Reviews, Volume 44, Issue 1, 31 October 2000, Pages 3-21
Saghir Akhtar, Marcus D. Hughes, Alim Khan, Mike Bibby, Majad Hussain, Qamar Nawaz, John Double and Pakeeza Sayyed
SummaryPlus | Full Text + Links | PDF (697 K)

6. **Cellular delivery of antisense oligonucleotides** • REVIEW ARTICLE
European Journal of Pharmaceutics and Biopharmaceutics, Volume 50, Issue 1, 3 July 2000, Pages 101-119
Irina Lebedeva, Lyuba Benimetskaya, C. A. Stein and Maria Vilenchik
SummaryPlus | Full Text + Links | PDF (605 K)

7. **Cutaneous vaccination: the skin as an immunologically active tissue and the challenge of antigen delivery** • ARTICLE
Journal of Controlled Release, Volume 66, Issues 2-3, 15 May 2000, Pages 199-214

Shawn Babiuk, Maria Baca-Estrada, Lorne A. Babiuk, Catherine Ewen and Marianna Foldvari
SummaryPlus | Full Text + Links | PDF (138 K)

8. ☐ **Application of membrane-based dendrimer/DNA complexes for solid phase transfection in vitro and in vivo** • ARTICLE
Biomaterials, Volume 21, Issue 9, May 2000, Pages 877-887
Anna U. Bielinska, Ann Yen, Huai Liang Wu, Kathleen M. Zahos, Rong Sun, Norman D. Weiner, James R. BakerJr. and Blake J. Roessler
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L5: Entry 25 of 26 File: USPT Apr 24, 2001

DOCUMENT-IDENTIFIER: US 6221959 B1
TITLE: Polynucleotide compositions

Brief Summary Text (17):
In accordance with the present invention, binding and solubilizing segments can be, independently of each other, linear polymers, randomly branched polymers, block copolymers, graft copolymers, star polymer, star block copolymer, dendrimers or have other architecture including but not limited to combinations of the above listed structures. For the purposes of the current invention all these structures are collectively called herein "block copolymers".

Brief Summary Text (56):
Additionally, cationic dendrimers, for example, polyamidoamines or polypropyleneimines of various generations (i.e., molecular weight), (Tomalia et al., Angew. Chem., Int. Ed. Engl. 1990, 29, 138) can be also used as polycation segments of block copolymers for gene delivery.

Brief Summary Text (79):
Filed concurrently with the parent of this application (Nov. 18, 1994) was Ser. No. 08/342,079 entitled "POLYMER LINKED BIOLOGICAL AGENTS". The entire disclosure of that application is incorporated herein by reference.

Brief Summary Text (104):
The ratio of the components of the polynucleotide composition is an important factor in optimizing the effective transmembrane permeability of the polynucleotides in the composition. This ratio can be identified as ratio .O slashed., which is the ratio of positively charged groups to negatively charged groups in the composition at physiological pH. If .O slashed.<1, the complex contains non-neutralized phosphate from the polynucleotide. The portions of the polynucleotides adjacent to the non-neutralized charges are believed to be a part of the shell of a polynucleotide complex. Correspondingly, if .O slashed.>1, the polycationic polymer or R-type segment will have non-neutralized charges, and the un-neutralized portions will fold so that they form a part of the shell of the complex. Generally, .O slashed. will vary from about 0 (where there are no cationic groups) to about 100, preferably .O slashed. will range between about 0.01 and about 50, more preferably, between about 0.1 and about 20. .O slashed. can be varied to increase the efficiency of transmembrane transport and, when the composition comprises polynucleotide complexes, to increase the stability of the complex. Variations in .O slashed. can also affect the biodistribution of the complex after administration to an animal. The optimal .O slashed. will depend on, among other things, (1) the context in which the polynucleotide composition is being used, (2) the specific polymers and oligonucleotides being used, (3) the cells or tissues targeted, and (4) the mode of administration.

Brief Summary Text (109):
The targeting molecules which can be associated with the polynucleotide compositions of the invention can also have a targeting group having affinity for a cellular site and a hydrophobic group. Such targeting molecules can provide for the site specific delivery and recognition in the body. The targeting molecule spontaneously associates with the polynucleotide complex and be "anchored" thereto through the hydrophobic group. These targeting adducts will typically comprise about 1% or less of the polymers in a final composition. In the targeting molecule, the hydrophobic group can be, among other things, a lipid group such as a fatty acyl group. Alternatively, it can be an ionic or nonionic homopolymer, copolymer, block copolymer, graft copolymer, dendrimer or another natural or synthetic polymer.

Detailed Description Text (47):
Male C57/B1/6 mice (weight: 20-24 g; obtained from the Russian Research Center of Molecular Diagnostics and Therapy, Moscow) received 50 .mu.l intravenous injections of Anti-HIV conjugate

or free Anti-HIV, at 0.18 OD.sub.260 /.mu.l dissolved in PBS. At defined times after the injections, blood sample were taken from the tail vein and the animals were sacrificed. The amount of radioactive material in blood or tissue sample was determined by liquid scintillation counting (after appropriate solubilizations). The results were as follows:

Detailed Description Text (147):
C. Caco-2 cells, originating from a human colorectal carcinoma (Fogh et al. J. Natl. Cancer Inst., 59-221-226, 1977) were kindly provided by Borchardt R. T. (The University of Kansas, Lawrence, Kans,). The cells were maintained in Dulbecco's Modified Eagle's Medium (DMEM), containing 10% heat-inactivated fetal bovine serum (FBS), 1% non-essential amino acids, benzylpenicilin (100 U/ml) and streptomycin (10 ug/ml), in an atmosphere of 90% air and 10% CO.sub.2 as described by Artursson (J. Pharm. Sci., 79:476-482, 1990). All tissue culture media were obtained from Gibco Life Technologies, Inc. (Grand Island, N.Y.). The cells were grown on collagen coated polycarbonate filter chamber inserts (Transwell, Costar Brand Tissue Culture Products, Contd.; pore size 0.4 um; diameter 24.5 mm). 250,000 cells were added to each insert and cells of passage number 32-45 were used. The cells were fed every second day and were allowed to grow and differentiate for up to 14 days before the monolayers were used in the following absorption experiments.

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